



भाकृअनुप - केन्द्रीय रोपण फसल अनुसंधान संस्थान
कासरगोड़, केरल, भारत
ICAR - CENTRAL PLANTATION CROPS, RESEARCH INSTITUTE
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F.No. 40(2)36-2020-Estate (Termination)

Dated: 09.06.2020

M/s Vishal Manpower & Security Consultants

#6, Kamala Towers, Near Urva market

Ashok Nagar P.O, Mangalore – 575 006

Sub: Termination of Job Contract work at CPCRI, Kasaragod – reg.

Ref: 1. This Office Tender Schedule F. No. 40(2)36-2019-Estate (C- Impr.) 40(2)36-2019-Estate (C- Prod.), 40(2)36-2019-Estate (C- Prot.), 40(2)36-2019-Estate (PB&PHT.), 40(2)36-2019-Estate (S.S.), dtd. 31.07.2019.
2. This Office Work Order F.No. 40(2)36-2019-Estate (C- Impr.) 40(2)36-2019-Estate (C- Prod.), 40(2)36-2019-Estate (C- Prot.), 40(2)36-2019-Estate (PB&PHT.), 40(2)36-2019-Estate (S.S.), dtd. 31.10.2019.

Sir,

With reference to the work order cited above, the Competent Authority has decided to terminate the Job contract works mentioned below, due to administrative exigencies and as per the terms & conditions appended in the tender document (Sl.No.18 & 30) by giving one month notice period. This letter may be treated as one month notice period for the termination of the contract. Accordingly, the contract shall stand terminated on completion of one month w.e.f the date of receipt of this letter.

Item No	Description of work	Actual Period of Work	Termination of job work w.e.f
I. Crop Improvement Division			
15	<p>Pollination Work</p> <p>The pollination work includes:</p> <ol style="list-style-type: none"> Noting dates of bunch opening, emasculation, bagging, pollination and removal of bag; recording number of female flowers pollinated. Emasculation (removal of male flowers from bunch) Bagging (covering bunch with a cloth bag three days before female flower receptivity) Collection of male spikes from dwarf and tall varieties Processing male flowers to extract pollen grains (sample of each batch of processed pollen to be provided to the lab for testing germination) Dusting pollens on the female flowers (on the bunch covered with bags; should be done before 11AM) Removal of bags (2-3 days after completing pollen dusting in a bunch) and tagging (tying bunches with tags showing bunch number and date of last pollination) Counting number of nuts set on the pollinated bunch <p>Work quantum indicators:</p> <ol style="list-style-type: none"> To complete pollination work in one bunch it may take 5-8 climbings One climber can attend to 50 palms Approximately 600 tall and 75 dwarf palms (6500 to 7000 bunches) <p>Conditions:</p> <ol style="list-style-type: none"> The climbers should be available with the department from 8AM to 5 PM and attend pollination related work as described above and as directed. All the day wise records of pollination should be maintained by individual climbers and provide as and when required. Work should be done as per the instructions. 	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020

[Handwritten Signature]

17	<p>Cleaning of glasswares</p> <ol style="list-style-type: none"> 1. Removal of media from culture tubes 2. conical flasks, Petri dishes, culture bottles 3. Soaking of culture tubes, conical flasks, Petri dishes, culture bottles in detergent solution (overnight) followed by brushing and cleaning. 4. Drying of glass wares in hot air oven <p>Note: 1000 test tubes 100 conical flasks 1000 Petri dishes 25 measuring cylinders 100 reagent bottle 100 small vials</p>	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020
18	<p>Lab work (Msc) for coconut tissue culture</p> <p>Preparation of stock solution of macro, micro elements, iron EDTA & Vitamins for MS, Y3 & other media for coconut.</p> <p>Preparation of hormone stock solution for Ms & Y3 media for coconut.</p> <p>Preparation of different media formulation & pouring into culture tubes / Petri dishes /conical flasks for tissue culture</p> <p>Preparation of inflorescence explants for culture initiation.</p> <p>Scoping of endosperm plug with embryo from coconut.</p> <p>Surface sterilization of endosperm plug, embryo & inflorescence</p> <p>Excision & inoculation of plumule from coconut embryo into different media combinations</p> <p>Inoculation of embryo or inflorescence into different media combination.</p> <p>Subculturing of cultures initiated from plumule, embryo or inflorescence into fresh media at 20 to 30 days interval.</p> <p>Documentation of culture details initiated at different periods.</p> <p>Cryopreservation studies of embryo and pollen</p> <p>Preparation of buffer/reagent solutions for molecular studies, Grinding of samples & extraction of DNA/RNA for clonal fidelity studies, Documentation of result & scoring of bands.</p>		
19	<p>Lab work (Msc) for arecanut tissue culture</p> <p>Preparation of stock solutions of macro, micro elements, iron EDTA and vitamins for MS, modified MS, Hoagland and Y3 and other media for Arecanut.</p> <p>Preparation of hormone stock solution for MS, Y3 and Hoagland media for Arecanut.</p> <p>Preparation of different media formulations and pouring into culture tubes /Petri dishes/conical flasks for tissue culture</p> <p>Surface sterilization of inflorescence collected from dwarf, hybrids and healthy arecanut palms from YLD hotspot area. (Season wise)</p> <p>Fine chopping of inflorescence and inoculation into different media combinations.</p> <p>Surface sterilization of embryo, excision of plumule from embryo or ovary from female flowers and inoculation into different media combinations</p> <p>Surface sterilization of seedling shoot column, fine chopping of meristematic tissues and inoculation into different media combinations.</p>		

	Subculturing of cultures into fresh media at 20 to 30 days interval. Documentation of culture details initiated at different periods.		
	Cryopreservation studies of embryogenic callus and pollen		
	Preparation of buffer/reagent solutions for molecular studies, Grinding of samples and extraction of DNA for clonal fidelity studies Documentation of results and scoring of bands		
II. Crop Production Division.			
5	Hill block- organic Farming	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020
	(a) Maintenance of plot, Planting of component crops which includes, bed preparation/pit opening and planting, input/fertilizer application, nut study. Work has to be carried out for 365 days.		
	(b) Lying of drip line, maintenance for drip irrigation. (* exact date of lying and maintenance is based on onset and cessation of monsoon).		
17	Maintenance of garden in the CPCRI premises (Main campus, Sagar, Kalpaka and Chandragiri guest houses premises which includes) Day to day maintenance and keep up of the garden at the CPCRI premises including lawn in the campus (removing the weds, watering and applying the fertilizers, pesticides etc. to the plants) Periodical moving the lawn with lawn mover – area 2500 sqm (approx.) Flower pots approximate 750 nos, cleaning watering and applying the fertilizer /pesticide etc De potting of flower pots and planting of flower plants as per the instructions (500 nos) Arranging and display of ornamental plants at office premises/ conference hall as and when required. Trimming of border plants periodically cutting, levelling, watering, applying fertilizers/ pesticides etc. (500 nos) Trimming of hedge plants like <i>phyllanthus</i> , <i>Durenta</i> , <i>Acalypha</i> (600 nos) Maintenance of garden type VI quarter (Director of CPCRI), including weeding, cleaning, watering etc. <u>NB: The work should be undertaken by gardener possessing at least two year's experience in the field of garden works.</u>		
III. PB & PHT Division			
1	Regular operation of SCADA in Open Top Chambers 2) Assisting in measurement of water potential, photosynthetic parameters, chlorophyll fluorescence of coconut & cocoa seedlings 3) Daily measurement of soil moisture using soil moisture probe to maintain plants at defined stress level 4) Assisting in estimation of shoot, root and total biomass in coconut seedlings under different moisture regime	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020
3	Assisting in biochemical analysis coconut and cocoa leaf samples at regular intervals (Approx. 100 samples each time). 1) Total phenol estimation, total and reducing sugars, MDA content, leaf chlorophyll content, Epicuticular wax content 2) Total soluble proteins along with protein profile under Poly-acrylamide gel electrophoresis (SDS-PAGE) and 2D gel electrophoresis		

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	<p>3) Antioxidant enzymes assay (SOD, Peroxidase, polyphenol oxidase, ascorbate peroxidase, carbonic anhydrase).</p> <p>4) Isozyme profile for SOD, peroxidase enzymes with Native PAGE</p> <p>5) Pollen collection and germination studies</p> <p>6) Assisting in Total DNA/RNA extraction from coconut leaves, cDNA preparation, qRT-PCR analysis</p>		
4	<p>Assisting in analysis of nutritional and biochemical constituents in coconut and arecanut (Approximately 100 samples each time)</p> <p>1) Analysis of coconut endosperm, testa, neera, etc. for various nutritional constituents like carbohydrates, lipids, proteins, etc.</p> <p>2) Total phenolics, flavonoids and antioxidant potential in various parts of coconut and arecanut.</p> <p>3) Proximate analysis of the unit operation based processed products of coconut like coconut milk, VCO, and other value added products</p>	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020
6	<p>Assisting in ice-cream unit</p> <p>1) Assisting in work related to the production of ice cream</p> <p>2) Processing of mature and tender nuts, cleaning the machineries etc.</p> <p>3) Washing the floor area, vessels and utensils</p> <p>4) Maintenance of production register</p> <p>Note: Unskilled</p>		
7	<p>Cleaning of glass wares, buckets & other utensils</p> <p>2) Drying in oven and staking in proper place (approx. 250 nos./day) in Biochemistry lab</p> <p>3) Grinding samples for biochemical analysis</p> <p>4) Irrigation of pots and cleaning of machineries in APC</p> <p>Note: Unskilled</p>		
IV. Crop Protection Division.			
1	<p>Production of <i>Trichoderma</i> formulations (Pathology)</p> <ul style="list-style-type: none"> • Assistance in carrying out molecular techniques like PCR, purification of DNA, amplified product, preparation of buffers etc.(weekly 50 samples) • Maintenance of <i>Trichoderma</i> nucleus cultures in the lab and sub-culturing regularly (once in 15 days). • Mass culturing of <i>Trichoderma</i> spp. for preparation of <i>Trichoderma</i> formulations viz. <i>Trichoderma</i> talc and <i>Trichoderma</i> coir pith cake.(100 litres of Trichoderm liquid culture per week) • Packing the talc powder in 2 kg capacity heat resistant polythene bags and sealing the bags. • Sterilization of the talc powder in autoclave. • Mixing the <i>Trichoderma</i> culture with talc powder. • Shade drying the formulation for 3-5 days. • Packing in polythene bags of required quantity and sealing the bags. (on an average 50 to 100 kg formulation per week based on the requirement) • Packing good quality coir pith in heat resistant polythene bags and 	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020

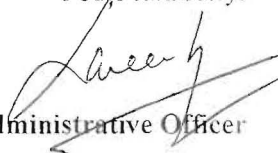
	<ul style="list-style-type: none"> sealing the bags. Sterilization of coir pith Mixing the <i>Trichoderma</i> culture with coir pith. Preparing the <i>Trichoderma</i> cake. Drying the cake in hot air oven, packing and sealing. Powdering and sterilization of Neem Cake. Mass production <i>Trichoderma</i> using sterilized neem cake substrate. 		
2	<p>Maintenance & Multiplication of insect cultures including parasitoids (<i>Barcon brevicornis</i> and <i>Goneozus nephantidis</i>) (Entomology)</p> <ul style="list-style-type: none"> Rearing of <i>Corcyra</i> larvae on broken wheat grains and transferring the larvae in to small test tubes for inoculation - 200 larvae/ day @30 paise/larvae Transferring emerged parasitoids in to bigger test tubes in aseptic condition and monitor the growth stage of the parasitoids includes; preparation of cotton plug and wax paper strips and fed the insects with honey - 1000-1200 parasitoids daily @ 30 paise per parasitoids Autoclaving used culture tubes, cleaning with soap solution drying and sterilizing in the oven - 200 small tubes 20 big tubes (weekly twice) Field collection of leaf eating caterpillar as and when incidence is reported and laboratory maintenance Field collection and maintenance of white grub cultures in the lab, cleaning and sterilizing of containers and filling with sterilized soil. Field collection of red palm weevil and rhinoceros beetles. Laboratory rearing of beetles by frequently providing coconut petioles. Collection of rhinoceros grubs and maintenance in laboratory Sterilization of laboratory equipments, preparation of fungal/ bacterial culture media for the multiplication of microbial biocontrol agents. 	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020
3	<p>Mass rearing of Greater wax moth, <i>Galleria mellonella</i> and mass production of entomopathogenic nematodes (Nematology)</p> <ul style="list-style-type: none"> Collection of honey combs and separation of <i>Galleria</i> larvae from bee hives maintaining by farmers at different places of Kasaragod. Preparation of ingredients (maize, wheat, bran etc.,) requirement of artificial diet its cleaning, drying and grinding process of desirable required standard and proper ratio for the multiplication of <i>Galleria</i> larvae. Preparation of artificial diet of 6 kg regularly at 20 days interval of 20 boxes for rearing different instar <i>Galleria</i> larvae for large scale production of EPN. This process will continue for throughout year. Collection of pupa and separation of adult moth of <i>Galleria</i> and placing in separate cages regularly at 15 days interval for hatching of eggs it is regular work to maintain required number larvae. Collection of eggs and keeping for larval hatching it is continuous process to avoid loss of culture. Assisting in selection and counting of fully grown <i>Galleria</i> larvae for EPN inoculation, collection EPN infested <i>Galleria</i> cadavers and its drying for 2 days, preparation of white trap chamber for placing cadavers to harvest the infective juveniles at 24 hours interval for regularly. Assisting in maintenance of hygiene in proper washing of all the devices using in multiplication of EPN at regular interval to avoid contamination Preparation of EPN storage process and assisting in packing, storing 		

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	and transportation of stored EPN vials/pouches to experiments filed on root grub management in YLD affected and non YLD plots at Karnataka and Kerala.		
	• Assisting in routine work of collection of soil samples and EPNs infected insect cadavers from trial plots and during survey work in different agro-climatic conditions of India for geomapping of EPN diversity.		
V. Social Science Division			
2	Carry out service works & maintenance and of statistics lab and Videoconferencing facility and for assisting in field surveys and data entry	01.11.2019 to 31.10.2020 (One Year) (Entire work)	01.07.2020

Please acknowledge the same.

Yours faithfully.


Chief Administrative Officer

Copy to:

1. The AHD, Crop Production Division and Chairman Contractual Service Committee. CPCRI, Kasaragod.
2. The AHD, Crop Improvement Division CPCRI, Kasaragod
3. The Scientists Crop Improvement, CPCRI, Kasaragod
4. The Technical Officer, Crop Improvement, CPCRI, Kasaragod.
5. The Sr. Fin. & Accounts Officer CPCRI, Kasaragod.
6. The DDO, CPCRI, Kasaragod.
7. The Asst. Labour Commissioner (Central). Office of the Regional Labour Commissioner, Kendriya Shram Sadan, Olimugal, By Pass Road, Kakkanad, Kochi-682 03
8. Website.
9. Guard file.